

Genetic diversity analysis of *Rhynocoris marginatus* Fabricius based on 18S Ribosomal RNA Gene (Heteroptera: Reduviidae)

Abstract

The *R. marginatus* are essential components of ecosystem, but also important in the biological control of insect pest, infesting a variety of agro ecosystem and medicine. The present investigation was carried out in the insect molecular genetic variation of 18S Ribosomal RNA gene from *R. marginatus*. The study was represent by the reduviid insect *R. marginatus* nucleotide gene sequences were translate amino acid sequence and obtained hydropathy, Domain, Transmembranes of proteins were calculated. The multiple gene sequence alignment of in-silico translated amino acid sequence of the partial ribosomal genes protein of *R. marginatus* were generated and the phylogenetic relationships were observed.

Keywords

R. marginatus, Nucleotide Sequence, Phylogeny, 18S RNA, Genetic Variation

Introduction

“Eukaryotic ribosomal DNA (rDNA) has several properties and was found useful for studying genetic variability and divergence within and between species”.¹“The Assassin bugs of the genus *Rhynocoris* are from diverse group of mostly insect pest with currently close to 190 species described worldwide and it have different morphs, biotypes, and ecotypes with various colours and shapes and well known for their role in bio control potential of the insect pests, yet their molecular relationships have not been established at molecular level”.^{2,3,4,5}“The typical insect mitochondrial genome is a circular, double stranded DNA molecule of about 12- 20 kb in length that contain 37 genes, 13 protein coding genes, 22 transfer RNAs (tRNA) and two ribosomal RNAs (rRNA)”.^{6,7}“Ribosomal RNA (rRNA) encoding genes (rDNA) and related genetic elements have been well studied for over six decades²⁴, with interests ranging from pharmaceutical and biochemical investigation to comparative biological studies garnering wealth of information on the structural, functional and evolutionary characteristics of these molecules. Phylogenetic studies in particular, have propagated a large number of rRNA gene sequences on public genetic databases, as the organismal universality and typically high gene copy number cell facilitate gene amplification and sequencing”.^{25,26}“Mitochondrial DNA has various interesting properties such as abundance in animal tissue, small size relatively simple genomic structure fast rate of evolution and a straight forward mode of transmission with a low level of recombination (due to its maternal inheritance). This makes it a valuable tool for comparative

genomic resolution".^{8,9,10} In this investigation was carried out based on available in ribosomal gene sequences from *R. marginatus* and amplified the partial nucleotide sequence of 18S ribosomal RNA gene.

Materials and methods

Collection of *R. marginatus*

A laboratory colony of *R. marginatus* were collected from Ayyanar Kovil Tropical Rain Forest bordering an agro ecosystem (altitude 389 MSL, latitude 76. 39° E and 10.45° N) near Rajapalayam, Virudhunagar District, Tamil Nadu, Southern India, during 2018-2021. The adults emerged were allowed to mate and the *R. marginatus* reared in the laboratory were used for experimental studies. Selected samples (n=5) were processed for DNA extraction following complete removal of ethanol. Total mtDNA was extracted from thoracic muscle or leg muscle of individual of the *R. marginatus* by phenol-chloroform method with minor modification as described by addition of 30 µl of proteinase k (20 mg/ml) and incubated for 16 hrs at 52°C.

Polymerase Chain Reaction, Sequencing and Analysis

The PCR was carried out to amplify the partial 18S ribosomal genes of 826 bp DNA fragment amplify form *R. marginatus*. It was amplified using two universal 18S gene specific primers: 18sf (5'-AAATTACCCACTCCCGGCA-3') and 18sr (5' TGGTGUGGGTTTCCCGTGTT-3'). The PCR products were separated on 2% agarose gel and visualized by ethidium bromide staining. The PCR products were purified using the HiYield PCR/ Gel extraction kit (RBC Biosciences, Taiwan) following the manufacturer's instructions. The purified amplicons were sequenced using the Big Dye Terminator Cycle sequencing ready reaction kit (Applied Biosystems Inc., USA) in the ABI prism 3100 Genetic analyzer. The sequencing of 18S amplicons from *R. marginatus* (n=5) was performed with the forward and reverse primer, and consensus sequence. Sequenced 18S gene of *R. marginatus* was assembled and analysed Editseq translate.

Results

We report here the isolation of the partial 18S ribosomal RNA gene sequence of the assassin bugs of *R. marginatus*. The 826 bp nucleotide sequence and conceptually translated aminoacid sequence of PCR amplicon of the 18S ribosomal RNA gene from *R. marginatus* (ure 1). The nucleotide composition of A+T percentage for the *R. marginatus* 18S gene is 53% and G+C percentage is 47%. The analysis into divulge the nucleotide frequencies of A-25%, T-28%, C-21% and G-26% (Table 1). In hydrophathy plot of the in-silicotranslated partial 18S gene protein of the 826 bp nucleotide sequence from *R. marginatus*. 18S gene protein was designates more of hydrophilic residues (mean by the peaks) and less of

hydrophobic residues (ure 2).

Molecular weight of the *R. marginatus* ribosomal 18S gene in 67641.30 μ and Residues 1-826, the average residues weight-81.890. Histogram plot of the in-silico translated nucleotide sequence of 18S gene indicates position from 1 to 826 bp, it reflect tiny residues and aliphatic, aromatic, non polar, polar residues and positive and negative residues of gene protein (ure 3).

In-silico translation with invertebrate mitochondrial genetic code in the Editseq translate revealed of 257 amino acid sequences for *R. marginatus*. The translation of the partial nucleotide sequence and its deduced amino acid sequences are shown in ure 1. Because of the codon preference, the A+T composition in *R. marginatus* is particularly biased at the second codon position, which totaled 16.57%. The A+T content at first and third positions are 16.09% and 15.85% respectively. The G+C composition at first and third are 15.72% and 15.37% respectively (Table 1). Multiple sequence alignment was carried out in the partial nucleotide sequence of ribosomal genes 16S, 18S, 28S from *R. marginatus*. And it used to help for investigation of codon similarity and divergence.

Genetic distances between the examined *R. marginatus* in the 18S gene have been generated by Neighbor-joining method. The minimum value of genetic distance among the examined 18S gene sequence from *R. marginatus* was 3.64 when compared with 16S, 18S respectively (Table 3). It was also observed that the percentage identity of 28S with 18S was 48.75% with a divergence of 2.64% (Table 3). The phylogeny was framework predicated on the aligned 18S gene sequences and 16S and 28S is a shown ure 4. The evolutionary history was inferred using the Neighbour joining method. The tree is drawn to scale, with branch lengths in the same units as those of the evolutionary distances used to infer the phylogenetic tree.

The phylogeny was created rested on the aligned 18s gene sequences and the tree obtained with the sum branch length of 0.33279. And here compined with three Ribosomal genes like 18S, 16S and 28S. The phylogenetic relationship revealed the existence of three clusters. The gene of the 16S formed one cluster and another 18S and 28S grouped forming other two clusters. The present study demonstrated the great effectiveness of mitochondrial 18S gene for inferring phylogenetic relationships at *R. marginatus* insect ribosomal gene level. Here reported to the phylogenetic relationship between the Ribosomal genes 16S, 18S and 28SRNA gene. (ure 4)

act cta ttg agg ccc cgt aat cgg aat aga gta cac ttt aaa tcc ttt aac aag gat cca

60

T L L R P R N R N R V H F K S F N K D P

20

| | |
|---------------------------------------------------------------------------------|-----|
| ttg gag ggc aag tct ggt gcc agc agc cgc ggt aat tcc agc tcc aat agc gta tat taa | 120 |
| L E G K S G A S S R G N S S S N S V Y - | 39 |
| agt tgt tgc ggt taa aaa gct cgt agt tgg ttc tgc gtc cca cgc tgt cgg ttc gcc gcc | 180 |
| S C C G - K A R S W F C V P R C R F A A | 58 |
| tgt cgg tgt aac tgg cat gtc gtg gca tgt cct gtc ggt ggt aaa cgg ggt ccc tgg tac | 240 |
| C R C N W H V V A C P V G G K R G P W Y | 78 |
| gac gta ggc ttt tat agc tga aat ctg tac cgt gtg tgt tcc cgt tta ccg atc tct cct | 300 |
| D V G F Y S - N L Y R V C S R L P I S P | 97 |
| act ccg gtg ctc tta aac gag tgt cga ggt agg ccg aca cgt tca ctt tga aca aat tag | 360 |
| T P V L L N E C R G R P T R S L - T N - | 115 |
| agt gct taa agc agg cta aaa tat ctg cct gaa tag tgg tgc atg gaa tga taa aac agg | 420 |
| S A - S R L K Y L P E - W C M E - - N R | 131 |
| acc tca gtt cta ttt tgt tgg ttt tag gaa tat gag gta atg atc aat gtg gac tgg cgg | 480 |
| T S V L F C W F - E Y E V M I N V D W R | 150 |
| ggg cat tcg tat tgc gac gtt aga ggt gaa att gtt gga tcg tcg caa gac gca cta gag | 540 |
| G H S Y C D V R G E I V G S S Q D A L E | 170 |
| cga aag cat ttg cca agt atg tct taa ttg atc aag aac gaa agt tag agg ttc gaa ggc | 600 |
| R K H L P S M S - L I K N E S - R F E G | 188 |
| gat cag ata ccg ccc tag ttc taa cca taa acg atg cca gcc agc gat ccg ccg atg ttc | 660 |
| D Q I P P - F - P - T M P A S D P P M F | 205 |
| gtt taa tga ctc ggc ggg gag ctt cta ctc ggg aaa cca aag ctt ttg ggt tcc ggg gga | 720 |
| V - - L G G E L L L G K P K L L G S G G | 223 |
| agt atg gtt gca aag ctg aaa ctt aaa gga att gac gga agg gca cca cca gga gtg gag | 780 |
| S M V A K L K L K G I D G R A P P G V E | 243 |
| cct gcg gct taa ttt gac tca cac ggg aaa ccc ccc cca aaa aaa a | 826 |
| P A A - F D S H G K P P P K K | 257 |

Figure 1: The 826 bp nucleotide sequence and conceptually translated amino acid sequences of PCR amplicon of the 18S ribosomal RNA gene from *R. marginatus*.

| Codon positions | A | T | C | G |
|------------------------|----------|----------|----------|----------|
| First position | 7.38 | 8.71 | 7.02 | 8.7 |
| Second position | 8.47 | 7.38 | 6.65 | 9.6 |
| Third position | 7.5* | 9.07* | 6.9 | 8.47 |

Table 1: Base Composition in the 826 bp nucleotide sequence of the 18S ribosomal RNA gene at the three codon positions in *R.marginatus*.

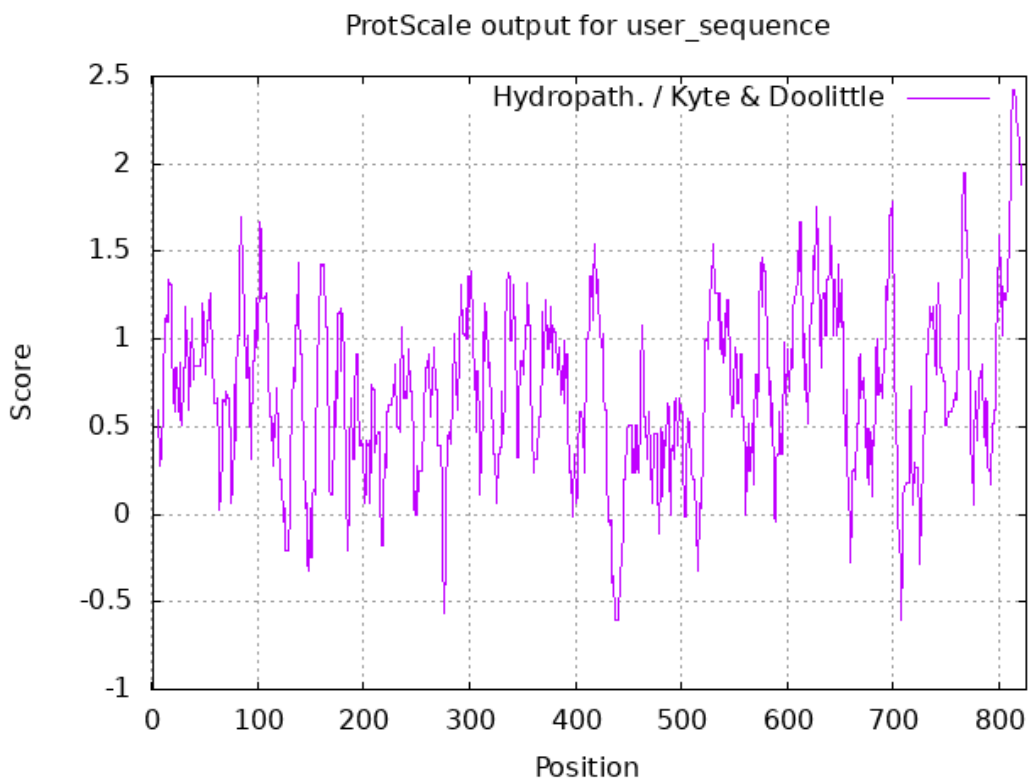


Figure 2: Hydropathy plot of the in-silico translated partial 18S ribosomal RNA gene protein from the 826 bp nucleotide sequence from *R. marginatus*

| No. | Nucleotide sequence obtained (bp) | A | A% | T | T% | C | C% | G | G% | AT% | GC % |
|-----|-----------------------------------|-----|-----|-----|-----|-----|-----|-----|-----|-----|------|
| 1 | 826 | 214 | 25% | 212 | 28% | 177 | 21% | 223 | 26% | 53% | 47% |

Table 2: Nucleotide composition of the partial sequenced 18S ribosomal gene from the *R. marginatus*.

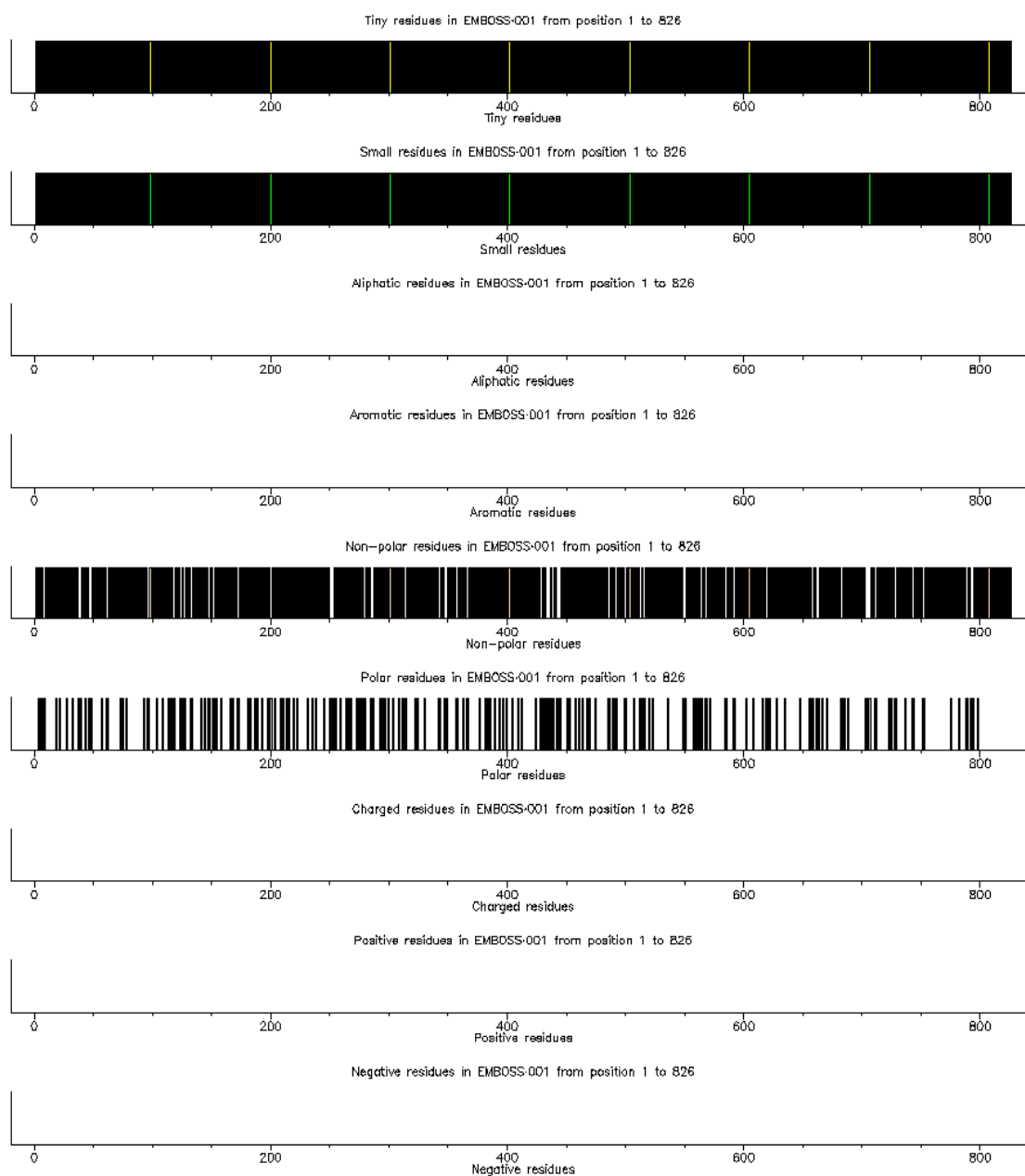
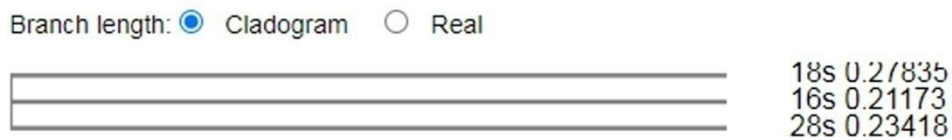


Figure 3: Histogram plot of the in-silico translated 826 bp nucleotide sequence of the 18S ribosomal RNA gene protein from *R. marginatus*.

Percentage identity

| | | 18s | 16s | 28s | | Gene name |
|------------|---|-----|-------|-------|---|-----------|
| Divergence | 1 | 18s | 50.99 | 48.75 | 1 | 18S |
| | 2 | 16s | 3.64 | 55.41 | 2 | 16S |
| | 3 | 28s | 2.64 | 2.18 | 3 | 28S |

Table 3: Percentage identity and divergence of the partial nucleotide sequence of the 18S, 16S and 28S Ribosomal RNA genes from *R. marginatus*



Tree Data

```
(
18s:0.27835,
16s:0.21173,
28s:0.23418);
```

Figure 4: Phylogenetic relationships of the three genes (16S, 18S, 28S ribosomal RNA gene) of *R. marginatus* based on nucleotide sequence of the PCR amplicon of the Ribosomal genes derived from Neighbor Joining Algorithm using Clustal Omega (Software 1.2.4)

Discussion

In the present investigation a 826 bp of the gene amplicons were recorded for the *R. marginatus* insect. On sequencing the 18S gene sequenced region matched with the already reported 18S gene sequence of some of the insect species that falls under the family of reduviidae. The sequence of the 18S gene generated in this study matched with sequence information results that are already reported in other insects Jon *et al.*, in

Hansenilla >1970 nucleotides¹¹ and Christane *et al.*, were studied about the assassin bugs in the same gene¹² and Yingqi *et al.*, also reported in the same gene in the assassin bug *Sigicoris stat*¹³, Uday kumar *et al.*, reported in *Linguataula serrata* insect¹⁴ and Gillespie *et al.*, were reported on *Apis melifera*¹⁵, Anil kumar *et al.*, on *Theileria annulata*¹⁶.

The nucleotide comparison and an amino acid sequences across the three ribosomal genes from *R. marginatus* indicated a higher divergence value of 3.64% and 2.64% in 18S and 16S genes respectively than that other 28S gene from *R. marginatus*. In related work has done and reported by already in the same insect. In higher genetic divergence values have been Cyt b and COI genes from four *Rhynocoris* species.
5

Analysis of the nucleotide sequence of the *R. marginatus* insect three Ribosomal genes are indicated higher nucleotide substitutions in 18S gene when compared to the other ribosomal genes 16S and 28S. Ambros *et al.*, were studied and reported into intrageneic phylogenetic relationships between thirteen species of *Coranus Curtis*² and Eisuke *et al.*, 2006 were reported the phylogenetic analysis of the insect order Odonata¹⁷, Mahendran *et al.*, were reported into *Bombycidae*.¹⁸ And already reported in other insects such as, *Chironomus* (Diptera) species Guryev *et al.*,¹⁹ and Jon *et al.*,¹¹, Yingqi *et al.*, in *Sigicoris*¹³, Austin *et al.*,²⁰, Arunkumar *et al.*,²¹ in *Bmbyxmori* and yogesh *et al.*,²² also reported in similar gene in various insect orders.

Here already reported to the phylogenetic analysis of various genes in species level. Such as Jon *et al.*, were reported *Hansenilla* has analysis phylogenetic tree and topological studies¹¹ and Christane *et al.*, 2009 were reported into Assassin bugs¹², Udhay kumar *et al.*, on *Linguatula serrata*¹⁴, Yingqi *et al.*, were reported in phylogenetic analysis in assassin bug *Sigicoris stat*¹³ and Gillespie *et al.*, 2006 were reported on *Apis melifera*¹⁵, Anil kumar *et al.*, 2022 on *Theileria annulata*.¹⁶ And some relative studies were reported in the same species such as Baskar *et al.*, were reported the phylogenetic relationships between the *Rhynocoris* species in four *Rhynocoris*, like *R. marginatus*, *R. longifrons*, *R. fuscipes* and *R. kumarri*.⁵ Ambrose *et al.*, reported in the genomic relationships among the four *harpatorine* reduviid species of *Rhynocoris*; like *R. kumarri*, *R. marginatus*, *R. Longifrons* and *R. fuscipes*.² In Eman *et al* were recently studied and reported on spiny bollworm were biologically controlled by using EPF as *Trichoderma aspereullum*. And its evaluate the phylogentic relationship between *Tricoderma* based on 18S rRNA partial sequence.²³

Conclusion

The results obtained not only have enriched our knowledge on biosystematics but have also supplemented multidisciplinary data. The results further reveals the utility of 18S ribosomal RNA sequence in multiple and phylogenetic analysis. In ribosomal 18S gene of *R. marginatus*, population which can be used to

develop molecular markers important for examining molecular genetic variation or gene diversity and understanding deep phylogenetic relationship the utility across the available heteropteran mtDNA ribosomal genome to facilitate informed gene choice for molecular study the *R. marginatus*. These results should allow the identification of the genetic variation and the analysis of phylogenetic information for understanding in *R. marginatus* genetic evolution.

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References

1. Fritz G. N., Conn J., Cockburn A. S and Eawright J. Sequence analysis of ribosomal DNA internal transcribed spaces 2 from populations of *Anopheles nunezyovari* (Diptera: Culicidae). *Molecular Biology and evolution*.1994; 11: 406-416.
2. Ambrose E., Arockia I. and Angeline K. D. Intrageneric phylogenetics based on mitochondrial DNA variation among fifteen harpactorine assassin bugs with four ecotypes and three morphs (Hemiptera : Reduviidae: Harpactorinae). *Zootaxa*. 2014;3779 (5): 540-550.
3. Putshkov V. G., Putskov P.V. A catalogue of assassin bugs genera of the world (Heteroptera : Reduviidae). *VINITI, Moskva*. 1985; 138.
4. Maldonado J. Systematic catalogue of the Reduviidae of the world (Insecta : Heteroptera). *Caribbean Journal of Science special editions, University of Puerto Rico, Mayaguez*. 1990; 694.
5. Baskar A., Dhanasekaran K. G., Thirumurugan A. R., Vignesh and Prakash. B. Molecular genetic variation and phylogeny of genus *Rhynocoris* based Mitochondrial Cytochrome c Oxidase subunit I gene (Heteroptera: Reduviidae). *Indian journal of Natural Sciences*. 2014; 4(22): 1-13.
6. Wolstenholme. Animal mitochondrial DNA structure and evolution. *Int Rev. cyto*.1992.
7. Yu Nie, Yu- Thian Fu. Yu Zhang Yun ping Deng, Wei wang, Ya Tu and Guo –Hau liu., 2021. Highly rearranged mitochondrial genome in falcoli perurisllice (Phthiroptera; Philoptera) from endangered eagles.
8. Avise J. C., Arnold J., Ball R. M., Bermingham E., Lamb T., Neigel J., Neigel J. E., Reeb C. A., Saunder N. C., 1987. Intraspecific phylogeograpy the mitochondrial DNA bridge between population genetics and systematic. *Annu, Rev.Ecol.syst*. 1987; 18: 489-522.
9. Moritz C., Dowling T. E., Brown W. M. Evolution of Animal mitochondrial DNA relevance for population biology and systematic. *Annu, Rev. Ecol. Syst*.1987; 269-292.
10. Arthur K., Maria K., Emeline I., Eric C., Julie P., Jerome C., Jerome M. Shot gun assembly of the

- assassin bug *Brontostoma colossus* mitochondrial genome (Heteroptera: Reduviidae). *Gene*. 2014; 4c: 39959-11.
11. Jon M. M., James R., Garey. and Jeffrey. W. S. Ecdysozoan phylogeny and Bayesian inference: First use of nearly complete 28s and 18s rRNA gene sequences to classify the arthropods and their kin. *Molecular phylogenetic and evolution*. 2003. 5: 213-216.
 12. Christians W., James B Munro. 2009. Molecular phylogeny of the assassin bugs (Hemiptera: Reduviidae), based on mitochondrial and nuclear ribosomal genes. *Molecular phylogenetics & Evolution*. 2009; 53, 287-299.
 13. Yingqi L., Hu L. and Wanzhi C. Revision of the Assassin Bug genus *Sigicoris* Stat Nov Based on morphological study and molecular phylogeny (Heteroptera: Reduviidae: Peiraatinae). *Insect-2022*; 13951.
 14. Uday kumar M. and Tasashi I. T. Molecular characterization and phylogeny of *Linguatata serrata* (Pentastomida : Linguatulidae) based on the nuclear 18s rDNA and mitochondrial cytochrome C oxidase I gene. *J. vet. Med . Sci*. 2017; 79 (2): 398-402.
 15. Gillespie, J. J., Johntoon, J. S., Cannones J. J., and Guttells R. R. Characteristics of the nuclear (18s, 5.8s, 28s and 5s) and mitochondrial (12s and 5s) rRNA genes of *Apis mellifera* (Insecta: Hymenoptera) structure, organization, and retro transposable elements. *Insect molecular biology*. 2006; 15 (5): 657-686.
 16. Anilkumar N., Ansu Kumari V. R., Kundave Sukhdeep V., Hira R. Molecular insights into the population structure and haplotype network of *Therila annulata* based on the small sub unit ribosomal RNA (18S rRNA) gene. *Infection, Genetics and Evolution*. 2022; 99: 105252.
 17. Eisuke H., and Eiti K. Phylogenetic analysis of the insect order odonata using 28s and 16s rDNA sequence; a comparison between data sets with different evolutionary rates. *Entomological science*. 2006; 9: 55-66.
 18. Mahendran B., Ghosh S. K and Kundu S. C. Molecular phylogeny of silk producing insects based on 16s ribosomal RNA and Cytochrome oxidase subunit I gene. Indian Academy of science. *J. Genet*. 2006; 85: 31-38.
 19. Guryev V., Makarevitch I., Blinov A. and Martin J. Phylogeny of genes Chironomous (Diptera) inferred from DNA sequences of mitochondrial Cytochrome b and Cytochrome oxidase I. *Mol. Phylogenet. Evol*. 2001; 19: 9-21.
 20. Austin J. W., Szalanski Allen L., and Kard Bradford M. Distribution and genetic variation of *Reticlatitermes* (Isoptera: Rhinotermitidae) in Oklahoma. *Florid Entomological Society*. 2004; 87(2):152-158.
 21. Arunkumar Muralidhar Melta K. P., Nagaraja J. Molecular Phylogeny of silkmoths reveals the origin of domesticated silkmoth, *Bombyx mori* from Chinese *Bombyx mandarina* and paternal

- inheritance of *Antheraea proylei* mitochondrial DNA. *Molecular phylogenetics and Evolution*. 2006; 40: 419-427.
22. Yogesh S., Shouche and Milind S patole. Sequence analysis of mitochondrial 16s ribosomal RNA gene fragment from seven mosquito species. *J. Bio Sci*. 2000; 25: 361-366.
 23. Eman M Abd-EIAzeem, Mohamed M Nada, Adel EA Amer, Rana HM Hussien. Isolation and identification of entomopathogenic fungi associated with the spiny bollworm and evaluation of their metabolites against the insect's biological parameters. *Egyptian journal of AgricultureResearch*. 2024; 102(1), 155-163.
 24. Paschallis N, Philip H , Irepan Savador-Martinoz, Maximillan J. Computational discovery of hidden breakes in 28s ribosomal RNAs across eukaryotes nd consequences for RNA integrity number scientific reports.2019; 9;19477/[https//doi.org,10.1038/41598-019-55573-1](https://doi.org,10.1038/41598-019-55573-1).
 25. Wosee, C. R., 1987; *Bacterial evolution microbiol.Rev*.51:221-271.
 26. Winker, S., and Woese C.R.,1991. A definition of the domains , Archaea, bacteria and eucarya, in terms of small subunit ribosomal RNA characteristics- syst. *Appl.Microbiol*. 14:305-310.